Review Article

Recent Advancement of Drug Delivery Through Microsphere

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ABSTRACT

Microspheres are multi-particulate drug delivery devices that are equipped to achieve a sustained or managed distribution of drugs to enhance bioavailability, stabilization and aim the drug at a fixed pace to a particular location. Depending on the consistency and form, biodegradable and nonbiodegradable polymers are used. There are all kinds of microspheres, such as bio-adhesive microspheres, magnetic microspheres, floating microspheres, toxic microspheres, chemical microspheres, biodegradable polymer microspheres, etc. The general characterization of microspheres depends on particle size, Electron Spectroscopy, Density determination, Isoelectric point, etc. The substance is stuck as a reservoir inside of the polymer in a microsphere or as a polymer-drug complex in a matrix. The opioid release sequence is responsible for an array of quantities. Most notably the factor that affects the release of the drug is the polymer type, its molecular weight, the excipients used in the formulation (surfactants, stabilizers, cross-linking agents), the copolymer type used, the association between drug polymers, etc. There are different approaches to administering a pharmaceutical drug in a prolonged controlled release mode to the target site.

Keyword: Microsphere, Polymeric Microsphere, Natural Polymers, Synthetic polymer, Alginates, Chitosan

INTRODUCTION

As per the team of research scientists, the healthcare industry is designing various initiatives to enhance clinical effectiveness and provide a controlled A well-designed response. and formulated pharmaceutical ingredients technology can change compliance and mitigate massive costs. In the range of 1-1000 micrometers, microspheres are spherical free-flowing powders. The descriptions of microspheres and microparticles are most often used together[1,2]. Microspheres, nevertheless, mean spherical void particles in a specific context. Any particle with a micrometer size spectrum between the provided polymer or protein-coated set is a microparticle. They are used uniquely for controlled or protracted release and recently as a selective delivery of medicines. In enhancing bioavailability and reducing the side effects of prescription drugs, the microsphere plays an important role[3,4]. The desired microsphere specifications are as follows-

- The capacity to integrate relatively high drug concentrations.
- Stability of the preparation with a clinically appropriate shelf life after synthesis.
- Regulated particle size and dispersibility in injection aqueous vehicles.
- Release of an active reagent over a large period with strong monitoring.

Benefits:

- They guard against unstable medications until after administration.
- Improving the drug's bioavailability.
- Provide continuous & consistent drug therapy activity.
- Decrease the frequency of doses & side effects.
- Provide enforcement with patients.
- Improving the solubility of poorly soluble drugs. Restrictions:
- Strong production prices.
- Bioactivity loss when making.
- Different rates of release from varying doses.
- Different GI physiological factors influence the release process.
- These may have potential toxicity if the release matrix has any flaw.
- You should not chew or smash these medication sources.
- Poor control of release rate.

Types Of Polymers In Microspheres

Depending on the consistency and form, the microsphere may be made with various types of polymers. But usually classified, according to the source, into two forms-

Synthetic polymers- Polymers that are synthesized in the laboratory by the polymerization of simple chemical molecules are known as synthetic polymers[5,6]. They are the following 2 types-

- A. Non-biodegradable polymers: These are natural polymers. E.g. Polymethyl methacrylate (PMMA), Acrolein, Glycidyl methacrylate, Epoxy polymers.[7]
- B. **Biodegradable polymers**: biodegradable polymers are unique polymers that break down into carbon dioxide, water, biomass, and other inorganic salts after a certain amount of time in the body. They should have

properties so they do not disintegrate, have controlled activity, and, most significantly, be non-toxic during the expected time. E.g. Glycosides, lactides & their copolymers, Poly alkyl cyanoacrylates, Poly anhydrides[8,9].

Natural polymers- For pharmaceutical applications, the use of natural polymers is beneficial because they are effective, readily accessible, and non-toxic. These are capable of chemical changes, theoretically biologically degradable, and, with a few exceptions, also biocompatible. Some representations of naturally obtained polymers used in pharmaceuticals are included in Table 1.0.

Table 1:

Source	Polymers	References
Plant Origin	Cellulose, Hemicellulose, Glucomannan, Agar, Starch, Pectin, Inulin,	[10]
	Rosin, Guar	
Animal Origin	Chitin, Alginates, Carrageenan Psyllium, Xanthan gum	[11]

Types Of Microspheres

Microsphere Bioadhesive

Bio-adhesive indicates adhesion to the surface of the biological membrane or mucus. Drug administration without invasion or discomfort is deemed to be safer. A more preferred route of administration is oral or administration.[12] The bio-adhesive nasal microsphere adheres to the abdominal, nasal, rectal, ocular, and respiratory mucus membranes to have improved communication and better therapeutic efficacy. These are beneficial in gene therapy and peptide distribution as well.[13]

Microsphere Magnetic

The super-molecular particles of the magnetic microsphere are small enough to circulate through capillaries without creating fewer than 4 micrometers of embolic occlusion.[14] They are ferromagnetic and are reported in micro-vessels with a 0.5-0.8 tesla magnetic field effect. The magnetic carrier here is magnetite, iron, nickel, cobalt, and Samarian cobalt which obtain the magnetic response from a magnetic field.[15]

Floating Microsphere

It is a way to reduce the drug's absorption for improved bioavailability. It increases the time of gastric residence, encouraging prolonged effects.[16] The bulk density of circulating microspheres is smaller than the gastric fluid and thus stays buoyant in the stomach without influencing the rate of gastric emptying. At the target pace, the rate of drug release steadily. This also reduces the likelihood of dumping impacts and doses.[17]

Radioactive Microsphere

There is a 10-30 nm radioactive microsphere that gets stuck in the capillary bed. Via intra-arterial radioembolization, radioactive microspheres accommodate liver meta-states. The radioactive microsphere includes a radioactive isotope that is pumped through the microvasculature via the artery, which also releases radiation through the tumors without influencing other tissues.[18]

Polymeric Microspheres

Two forms of polymeric microspheres are—

- A- Biodegradable Polymeric Microsphere- With the premise that they are ultimately biodegradable, biocompatible, and even bioadhesive, conventional polymers like starch are included. Along with its high degree of aqueous media swelling property, biodegradable polymers prolong the residence time as gel formation results in contact with the mucous membrane. The rate and degree of drug release are sustainably controlled by the concentration of polymers and the release pattern.[18]
- B- Synthetic Polymeric Microsphere- In addition to becoming frequently used as bulking agents, fillers, embolic particles, drug delivery systems, etc., synthetic polymeric microspheres are generally used in medicinal applications and have proved to be secure and biocompatible,

but the key downside of these microspheres is because they seem to migrate away from the injection site and lead to and contribute to possible harm, embolism and further damage to the tissue.[19]

Method Of Preparation Of Microspheres Single emulsion method

This method is one of the oldest methods for microsphere preparation. In this method, the natural polymers are dispersed in an aqueous medium along with a drug with continuous agitation followed by dispersion in a non-aqueous medium e.g. oil. In the next step, the formed globules are cross-linked by heat or chemicals. Chemicals include glutaraldehyde, diacid chloride, formaldehyde, etc. However, it has the limitation that this technique in case of hydrophilic drugs causes burst release of the drug upon administration due to the accumulation of the drug crystals on the surface of microspheres due to the partitioning of the drug in the aqueous phase of the emulsion [20,21].

Double emulsion technique

This method includes the formation of multiple emulsions or double emulsion. Peptides, proteins, and water-soluble drugs are appropriate candidates for this method. Both natural and synthetic polymers can be used for this technique. In this method, the aqueous solution is dispersed in an organic phase which consists of the polymer. These polymers entrap the aqueous phase consisting of the active ingredient. This primary emulsion is then subjected to homogenization or sonication and finally into poly-vinyl alcohol. This results in a double emulsion.[22] Now the final step is the removal of the solvents usually done by solvent extraction and solvent evaporation and finally the addition of a large amount of water. Filtration is done and excess oil is removed by washing with n-hexane, acetone, or any other organic solvent. A variety of hydrophilic drugs such as luteinizing hormone-releasing hormone (LH-RH) agonists, vaccines, proteins/peptides, and traditional molecules are successfully introduced into the microsphere using the evaporation/extraction dual emulsion solvent process.[23]

Phase separation and coacervation

In this technique, the polymer is dissolved in a suitable solvent along with a drug to make an aqueous solution in case the drug is hydrophobic or into the polymer itself. Phase separation is accomplished by a change in PH, the addition of salt, the addition of incompatible polymer, etc. This method is particularly for peptides, proteins, etc. which are water-soluble and also for hydrophobic e.g. Steroids[24,25].

Spray drying and spray congealing

The drug is dispersed at high-speed homogenization into a polymer which is previously dissolved in an organic solvent like acetone or dichloromethane. By using the cyclone separator, microparticles are separated from the hot air while the solvent residue is eliminated by vacuum-drying. One of them the key advantages of this process is practical viability under aseptic conditions.[26]

Solvent extraction

Here, removal of the organic phase is done by extraction of the organic solvent. The method involves water-miscible organic solvents such as isopropanol. The organic phase is removed by extraction with water. This process decreases the hardening time for the microspheres. The process involves the direct addition of the drug or protein to polymer organic solution. The rate of solvent removal by extraction method depends on the temperature of the water, a ratio of emulsion volume to the water, and the solubility profile of the polymer.[27]

Emulsion solvent evaporation

Initially, the polymer is dissolved in acetone followed by the addition of drug into the polymer solution, then by the addition of magnesium stearate. Then this dispersion was added to a mixture containing liquid paraffin while stirring on a mechanical stirrer. Stirring was continued unless acetone evaporated. The microspheres formed are filtered and washed. [28]

lonic gelation technique

lonotropic gelation is based on the ability of polyelectrolytes to cross-connect to form hydrogel beads often called gel spheres in the presence of counter ions.[29] Gel spheres are spherical crosslinked polymeric hydrophilic entities capable of substantial gelation and swelling of synthetic biological fluids and the release of the substance regulated by polymer relaxation. Polymers used in formulations are as follows-

Alginates, Gellant gum, Chitosan, Pectin, and Carboxymethyl cellulose are widely used for the encapsulation of drugs by this technique.

(a) Alginates

Alginates or alginic acid are Obtained from brown seaweed usually white to yellowish-brown. Sodium alginate when comes in contact with polyvalent ions such as calcium, barium, or strontium it forms a cross-linking. Sodium alginate is a sodium salt of alginic acid a natural polysaccharide and a linear polymer composed of 1,4-linked [3-D Mannuronic acid (M) and a D-guluronic acid (G) residues in varying proportions and arrangement. An insoluble salt is formed by the complex formation between sodium ions from guluronic acid and calcium ions. Alginate possesses mucoadhesive properties which could increase the contact time between microcapsules and absorptive sites, and therefore, could enhance the uptake of encapsulated drugs.[30]

(b) Gellan

Gellan gum is a bacterial ex O-polysaccharide prepared by fermentation of *Sphingomones Elodea*. It is dissolved in water at high temperatures. When the temperature is decreased, the chains undergo a conformational transition from random coils to double helices. Then rearrangement of a double helix occurs leading to the formation of the gel[33].

(C) Chitosan

Chitosan is natural poly(amino saccharide), having structural characteristics similar to glycosaminoglycan, is non-toxic, and easily bioabsorbable. Chitosan due to its antacid and antiulcer characteristics prevents or weakens drug irritation in the stomach. It could be used for the preparation of various polyelectrolyte complex products with natural polyanions such as xanthan, alginate, and carrageenan. Among these, complexes, the chitosan alginate complex may be the most important drug delivery hydrogel system. Chitosan has properties like it is a weak base, insoluble in water and organic solvents, but, it is soluble in dilute aqueous acidic solution (pH < 6.5), which can convert the glucosamine units into a soluble form of protonated amine (R_NH3+). Chitosan gets precipitated in alkaline solutions or with polyanions and forms a gel at a lower pH[31].

Emulsification heat stabilizing method

The drug is dissolved in albumin solution and polymer in acetic acid. A primary emulsion is made from it by dissolving the aqueous phase into the oil phase. This primary emulsion is dispersed in the oil phase which is previously heated 70°c using a magnetic stirrer at 1000 rpm. The excess oil is removed by washing it with ether. The obtained microspheres are dried in a vacuum and stored in a desiccator.[32]

General Characterization Of Microspheres

Characterization or evaluation of the microsphere is one of the most important analyses to formulate any kind of microsphere. There are various parameter which helps to analyze the physical as well as chemical properties of the drug-loaded microsphere.[35] The following fig no 1.0 contains various parameters to check the efficacy of the formulated microsphere by using different techniques. [36,70]



Fig No 1.0- Various Evaluation Parameters

Factors Affecting Drug Release Kinetics

The drug is entrapped within a microsphere as a reservoir within a polymer or in a matrix as a polymer-drug complex. An array of factors is responsible for the drug release pattern. Most important being the type of polymer, its molecular weight, excipients used in the formulation (surfactants, stabilizers, cross-linking agents), the type of copolymer used, drug-polymer interaction, etc. Also, the size of the microsphere has a significant effect on drug delivery. Usually, size and drug delivery follow an inverse relationship. The degradation of the polymer takes place in two ways that are bulk eroding and surface eroding. Poly anhydride is hydrophobic monomers with kinetically unstable bonds that prevent water penetration into the bulk and degrade as its surface erodes. While PLG readily allows water penetration and degradation. In bulk erosion, the loading dose is released followed by slow and controlled release. Bulk erosion can occur by two mechanisms i.e. pore diffusion and osmotically by driven diffusion Drug release from non-biodegradable polymers generally depend on whether they are matrix or reservoir type[37,38].

The molecular weight can affect the release rate of the drug and it is self-explanatory that an increase in molecular weight would decrease the drug release. It holds for small molecules, peptides, etc. However molecular weight has little effect on the release rate of polyanhydride.[39] All types of microspheres prepared with different molecular weights PLA and PLGA displayed a high protein-loading efficiency (> 80%) but their size was strongly influenced by polymer molecular weight (3000 versus 100,000). Protein release pattern was influenced by both polymer molecular weight and composition (PLA versus PLGA). The release rate was lower from PLA microspheres than from PLGA microspheres. In contrast, a continuously increasing release rate preceded by a burst was observed for low molecular weight (3000) PLGA microspheres.[40]

Approaches For Disease Targeting Through Microspheres

There are some benefits of the use of microspheres as a drug delivery system, such as improved efficiency and decreased toxicity of the inserted agents of non-targeted cells and tissues. There are also drawbacks, however: after several weeks, microspheres are denatured and are also fairly weak and therefore not conveniently mass-produced. [41] Through lodging them in the final organ vessels, biodegradable microspheres may be used to pass drugs to organs. Its effectiveness depends on the scale of the microsphere used and the (intravenous / intra-arterial) mode of administration. New treatments, of neoplasia targeting and giving high priority, are continuously being explored and developed. For both kinds of procedures, the popular factor is the need for targeting to prevent opioid side effects. [42,43] In microsphere Selective Internal Radiation Therapy (SIRT) is emerging as a potential treatment modality via hepatic arterial administration in the care of patients with primary and metastatic liver cancer [44-47]. The selectivity of the operation is due to the particular hepatic arterial flow pattern that comprises the vast majority of the supply of blood to the tumor. [48,49].

Enhanced and targeted drug delivery using biodegradable microspheres is emerging as a

promising approach for cancer therapy. The main objective of the present research was to formulate, characterize, and evaluate iron oxide (magnetic) containing a bovine serum albumin-based microsphere drug delivery system, capable of efficiently delivering sulforaphane, a histone deacetylase inhibitor, for an extended period in vivo. Magnetic microspheres were prepared by spraydrying and characterized for their physicochemical properties and dissolution profile. [50]

Recently, multiple particle engineering developments have arisen that have made it possible for inhaled microspheres to attempt to control pulmonary biopharmaceuticals and enhance clinical effectiveness for local and systemic therapies. These microspheres may be configured to sustain drug release, extend lung retention, accomplish drug targeting and/or improve drug absorption, and thereby investigate the ability to minimize the duration of dosing and/or the dosage of medicines while retaining clinical effectiveness and/or reducing adverse effects. [51]

Drug	Site of Action	Disease Targeting	Reference
90Y-microsphere therapy	Injected into the hepatic artery	Three-dimensional tumour dosimetry for hepatic	[52]
Yttrium-90 (90 Y)	Single whole-liver injection or single session selective bilobar administration. Unilobar disease was treated with a selective lobar injection.	Unresectable metastatic liver neuroendocrine tumors (NET).	[53]
Yttrium-90resin microspheres	Intra-arterial	Hepatocellular carcinoma (HCC)	[54]
Yttrium-90 ((90)Y) (Microsphere radioembolization)	Intra-arterial	liver malignancies	[55]
Yttrium-90 microspheres (SIR-Spheres and Thera Spheres)	Injected via the hepatic artery	Liver cancer	[56,57]
Cytotoxin-loaded microspheres	Delivered through a catheter, surgically implanted directly into either the subclavian artery or into a branch of the subclavian artery, usually the thyrocervical trunk.	Breast cancer	[58-60]
Small quantity of anti- neoplastic agent to the inner surface of the colon (5-fluorouracil). Guar gum microspheres are a potential system	Deliver drugs to tissues, and not through tissues.	Colorectal cancer	[61,62]

Table 2: The below table summarizes the different approaches used for testing microsphere activity

delivery of methotrexate to the colon, for chemotherapy of colorectal cancer.			
Paclitaxel-loaded PLGA microspheres. Conjugating camptothecin onto PEGylated microspheres prolongs the release of camptothecin in-vitro and enhances the anti-cancer efficacy.	Effective vehicles for pulmonary chemotherapeutic drug delivery	Lewis lung carcinoma cells	[63,64]
cisplatin prepared as L- Lactic acid and glycolic acid copolymer microspheres	Intra-peritoneal	Ovarian cancer	[65,66]
Paclitaxel	Intra-articular	Arthritis	[67]
Sulforaphane	Intraperitoneally	B16 melanoma cells	[68]
Azithromycin 2.0g microspheres	Single-dose azithromycin administration	Lower respiratory tract infections.	[69]

Applications of Microspheres

Drug loaded microsphere involve targeting via different delivery like topical drug delivery, targeting

drug delivery including this it has various applications as shown in the fig No 2.0. [70,71,72]



Fig .2: Applications of Microsphere

CONCLUSION

The microsphere is a small spherical entity with diameters ranging from 1 μ m to 1000 μ m in the

micrometer scale. Microspheres are characteristically free-flowing powders composed of naturally biodegradable proteins or synthetic polymers that preferably have a particle size of fewer than 200 µm. There are different approaches to administering a pharmaceutical drug in a prolonged controlled release mode to the target site. Using microspheres as carriers for drugs is one such strategy. Popular examples of floating structures are floating tablets and floating microspheres. The administration of controlled-release medications employs drugencapsulating systems from which therapeutic agents can be extracted over lengthy periods at controlled speeds, spanning from days to months. These technologies provide several benefits over conventional drug delivery approaches, including medication release rate tailoring, delicate drug safety, and improved patient satisfaction and compliance. Thanks to their potential to encapsulate a range of medications, biocompatibility, good bioavailability and continuous drug release characteristics, polymeric microspheres are perfect controlled vehicles for many distribution applications.

REFERENCE

- Sahil, K., Akanksha, M., Premjeet, S., Bilandi, A., & Kapoor, B. Microsphere: A review. Int. J. Res. Pharm. Chem, 2011;1;1184-98.
- 2. Mahale, M. M., & Saudagar, R. B. Microsphere: A Review. J. Drug Deliv and Ther, 2019;9;854-56.
- Kutmalge, M. D., Ratnaparkhi, M. P., Wattamwar, M. M., & Chaudhari, S. P. (2014). A REVIEW ON MICROSPHERE. *Pharma Science Monitor*, 5.
- 4. Hire, N. N., & Derle, D. V. Microsphere as drug carrier: a review. Int. J. Adv. Res, 2014;2;901-13.
- Calori, I. R., Braga, G., de Jesus, P. D. C. C., Bi, H., & Tedesco, A. C. Polymer scaffolds as drug delivery systems. *Eur. Polym. J*,2020; 109621.
- 6. Abasian, P., Ghanavati, S., Rahebi, S., Nouri Khorasani, S., & Khalili, S. Polymeric nanocarriers in targeted drug delivery systems: A review. *Polym Adv Technol*,2020.
- Bhosale, R. R., Gangadharappa, H. V., Osmani, R. A. M., & Gowda, D. V. Design and development of polymethylmethacrylate-grafted gellan gum (PMMAg-GG)-based pH-sensitive novel drug delivery system for antidiabetic therapy. *Drug Deliv Transl Res*, 2020;1-17.
- Stewart, S. A., Domínguez-Robles, J., McIlorum, V. J., Mancuso, E., Lamprou, D. A., Donnelly, R. F., & Larrañeta, E. Development of a biodegradable subcutaneous implant for prolonged drug delivery using 3D printing. *Pharmaceutics*, 2020;12;105.
- Choubdar, N., & Avizheh, S. (2020). Nanotechnology Based Delivery Systems of Drugs Currently Used to Treat Alzheimer's Disease. Nanosci Nanotechnol-Asia, 2020; 10;228-47.

- Alves, T. F., Morsink, M., Batain, F., Chaud, M. V., Almeida, T., Fernandes, D. A., ... & Severino, P. Applications of Natural, Semi-Synthetic, and Synthetic Polymers in Cosmetic Formulations. *Cosmetics*, 2020;7;75.
- Tong, X., Pan, W., Su, T., Zhang, M., Dong, W., & Qi, X. Recent advances in natural polymer-based drug delivery systems. *React Funct Polym*, 2020;148;104501.
- Rahaman, S. T., & Mukherjee, J. A REVIEW ON MUCOADHESIVE MICROSPHERES AS AN EFFICIENT DRUG DELIVERY SYSTEM.
- Beg, S., Rahman, M., Panda, S. K., Alharbi, K. S., Alruwaili, N. K., Singh, P. K., ... & Singh, B. (2020). Nasal Mucoadhesive Microspheres of Lercanidipine with Improved Systemic Bioavailability and Antihypertensive Activity. J Pharm Innov, 2020;1-10.
- 14. Wang, S., Pi, L., Wen, H., Yu, H., & Yang, X. Evaluation of novel magnetic targeting microspheres loading adriamycin based on carboxymethyl chitosan. J Drug Deliv Sci Technol, 2020;55;101388.
- 15. Gurung, B. D., & Kakar, S. An overview on microspheres. Int J H Clin Res, 2020:3;11-24.
- Kumal, V. B., Thapa, C., Ghimire, P., Chaudhari, P., & Yadhav, J. Formulation and optimization of Enalapril Maleate-loaded floating microsphere using Box–Behnken design: in vitro study. J Appl Pharm Sci,2020;10; 095-104.
- Ali, A., Pillai, H. H., Mathew, P., Beena, P., Das, C., & Abraham, E. Formulation and Evaluation of Floating mucoadhesive Microspheres loaded with Antiulcer drug. Res J Pharm Technol, 2020;13;3759-64.
- Arranja, A. G., Hennink, W. E., Chassagne, C., Denkova, A. G., & Nijsen, J. F. W. Preparation and characterization of inorganic radioactive holmium-166 microspheres for internal radionuclide therapy. *Mater Sci Eng;: C*, 2020;106, 110244.
- Zhong, H., Gao, X., Qiu, Z., Sun, B., Huang, W., & Li, J. Insight into β-cyclodextrin polymer microsphere as a potential filtration reducer in water-based drilling fluids for high temperature application. *Carbohydr Polym*, 2020;249;116833.
- 20. Wei, Y., Wu, Y., Wen, K., Bazybek, N., & Ma, G. Recent research and development of local anesthetic-loaded microspheres. J Mater Chem B, 2020;8:6322-32.
- 21. Gurung, B. D., & Kakar, S. An overview on microspheres. Intern J H Clin Res, 2020;3;11-24.
- 22. Yang, Y. Y., Chung, T. S., & Ng, N. P. Morphology, drug distribution, and in vitro release profiles of biodegradable polymeric microspheres containing protein fabricated by double-emulsion solvent extraction/evaporation method. *Biomaterials*, 2001;22;231-41.

- 23. Du, L., Cheng, J., Chi, Q., Qie, J., Liu, Y., & Mei, X. Biodegradable PLGA microspheres as a sustained release system for a new luteinizing hormonereleasing hormone (LHRH) antagonist. *Chem pharmal bulle*, 2006;54;1259-65.
- 24. Bayomi, M. A., Al-Suwayeh, S. A., El-Helw, A. M., & Mesnad, A. F. Preparation of casein–chitosan microspheres containing diltiazem hydrochloride by an aqueous coacervation technique. *Pharm Act Helv*, 1998;73;187-92.
- 25. Sunitha, S., Amareshwar, P., Santhosh Kumar, M., & Chakravarti, P. Preparation and evaluation of tramadol hydrochloride microspheres by coacervation phase separation technique using various solvents and non-solvents. J Glob Pharma Technol, 2011;3;33-41.
- Cordeiro, P. A. U. L. A., Temtem, M., & Winters, C. O. N. R. A. D. Spray congealing: applications in the pharmaceutical industry. *Chem. Today*, 2013;31;69-72.
- 27. Bitz, C., & Doelker, E. Influence of the preparation method on residual solvents in biodegradable microspheres. *Int j pharm*, 1996;131;171-81.
- 28. Kim, B. K., Hwang, S. J., Park, J. B., & Park, H. J. Preparation and characterization of drug-loaded polymethacrylate microspheres by an emulsion solvent evaporation method. *J microen*, 2002;19; 811-22.
- 29. Sahu, S., Chourasia, A., Toppo, A., & Asati, A. (2012). Formulation and evaluation of captropril microspheres by ionic gelation technique. *Int j pharm life sci*,2012; *3*.
- Das, M. K., & Senapati, P. C. Evaluation of furosemide-loaded alginate microspheres prepared by ionotropic external gelation technique. *Acta Pol Pharm*, 2007;7; 253-62.
- Sivakumar, M., & Rao, K. P. Preparation, characterization and in vitro release of gentamicin from coralline hydroxyapatite-gelatin composite microspheres. *Biomaterials*, 2002;23;3175-81.
- 32. Elzoghby, A. O., Samy, W. M., & Elgindy, N. A. Protein-based nanocarriers as promising drug and gene delivery systems. *J* controlled release, 2012;161;38-49.
- 33. Jampen, S., Britt, I. J., & Tung, M. A. Gellan polymer solution properties: dilute and concentrated regimes. *Food Res Int*, 2000;33;579-86.
- Bodmeier, R., & Chen, H. Preparation and characterization of microspheres containing the anti-inflammatory agents, indomethacin, ibuprofen, and ketoprofen. J controlled release, 1989;10;167-75.
- 35. Benita, S., Benoit, J. P., Puisieux, F., & Thies, C. Characterization of drug-loaded poly (d, l-lactide) microspheres. *J Pharm Sci*, 1984;73;1721-24.

- Yamamoto, T., Sugimoto, T., Suzuki, T., Mukai, S. R., & Tamon, H. Preparation and characterization of carbon cryogel microspheres. *Carbon*, 20020;40;1345-51.
- Freiberg, S., & Zhu, X. X. Polymer microspheres for controlled drug release. *Int | pharm*, 2004;282; 1-18.
- Berkland, C., Kipper, M. J., Narasimhan, B., Kim, K. K., & Pack, D. W. Microsphere size, precipitation kinetics and drug distribution control drug release from biodegradable polyanhydride microspheres. *J Controlled Release*, 2004;94;129-41.
- 39. Wu, J., Kong, T., Yeung, K. W. K., Shum, H. C., Cheung, K. M. C., Wang, L., & To, M. K. T. Fabrication and characterization of monodisperse PLGA-alginate core-shell microspheres with monodisperse size and homogeneous shells for controlled drug release. Acta biomaterialia, 2013;9;7410-19.
- 40. Zolnik, B. S., Leary, P. E., & Burgess, D. J. (2006). Elevated temperature accelerated release testing of PLGA microspheres. J Controlled Release, 2006;112;293-300.
- 41. Suzuki K. Activated CD4+ T cells preferentially take up lipid microspheres, but resting cells do not. Clin Exp Immunol 1994;99:479-85
- 42. Jayaprakash S, Halith SM, Firthose PUM, Kulaturanpillai K, Abhijith, Nagarajan M. Preparation and evaluation of biodegradable microspheres of methotrexate. Asian J Pharm 2009;3:26-9.
- 43. The Internet Encyclopaedia of Science. Available from: http:// www.daviddarling.info/encyclopedia/M/microsphere .html. [last cited on 2009 Jul 29]
- Gray BN, Anderson JE, Burton MA, Van Hazel G, Codde J, Morgan C, Klemp P: Regression of liver metastases following treatment with yttrium-90 microspheres. Aust N Z J Surg 1992, 62:105-10.
- 45. Lau WY, Leung WT, Ho S, Leung NWY, Chan M, Lin J, Metreweli C, Johnson P, Li AKC: Treatment of inoperable hepatocellular carcinoma with intrahepatic arterial yttrium-90 microspheres: a phase I and II study. Br J Cancer 1994, 70:994-99.
- Stubbs RS, Cannan RJ, Mitchell AW: Selective internal radiation therapy (SIRT) with 90Yttrium microspheres for extensive colorectal liver metastases. Hepato-gastroenterology 2001, 48:333-37.
- 47. Salem R, Thurston KG, Carr Bl, Goin JE, Geschwind JFH: Yttrium90 microspheres: Radiation therapy for unresectable liver cancer. J Vasc Interv Radio 2002, 13:S223-29.
- Kennedy AS, Coldwell D, Nutting C, Murthy R, Wertman DE, Loehr SP, Overton C, Meranze S, Niedzwiecki J, Sailer S: Resin Y-90- microsphere brachytherapy for unresectable colorectal liver

metastases: Modern USA experience. Int J Radiat Oncol Biol Phys 2006, 65:412-425.

- 49. Gulec SA, Fong Y: Y-90 microsphere selective internal radiation treatment of hepatic colorectal metastases. Arch Surg in press.
- Do, D., D'Souza, Enriquez, & Rizvi. Formulation and evaluation of drug-loaded targeted magnetic microspheres for cancer therapy. Int J Nanomedicine, 2013;1393.
- Weers JG, Bell J, Chan H-K, Cipolla D, Dunbar C, Hickey AJ, Smith IJ. Pulmonary formulations: What remains to be done? Journal of Aerosol Medi Pulm Drug Del. 2010;23:S-5–S-23.
- 52. P L Roberson, R K Ten Haken, D L McShan, P E McKeever, W D Ensminger. Three-dimensional tumor dosimetry for hepatic yttrium-90microsphere therapy. J Nucl Med. 1992 May;33;735-8.
- 53. Jia, Z., Paz-Fumagalli, R., Frey, G., Sella, D. M., McKinney, J. M., & Wang, W. Single-institution experience of radioembolization with yttrium-90 microspheres for unresectable metastatic neuroendocrine liver tumors. J Gastroen Hepatology, 32; 1617–23.
- 54. Wang, E. A., Stein, J. P., Bellavia, R. J., & Broadwell, S. R. Treatment options for unresectable HCC with a focus on SIRT with Yttrium-90 resin microspheres. International Journal of Clinical Practice, Int J Clin Pract. 2017 Nov;71.
- 55. W Bult, M A D Vente, B A Zonnenberg, A D Van Het Schip, J F M Nijsen. Microsphere radioembolization of liver malignancies: current developments. Q J Nucl Med Mol Imaging. 2009 Jun;53;325-35.
- 56. Sharma RA, Guy A, Hazel V, Morgan B, Berry DP, Blanshard K, et al. Radioembolization of liver metastases from colorectal cancer using yttrium-90 microspheres with concomitant systemic oxaliplatin, fluorouracil, and leucovorin chemotherapy. J Clin Oncol 2007;25:1099-106.
- 57. Murthy R, Habbul A. Trans-arterial hepatic radioembolisation of yttrium-90 microspheres. Biomed Imag Interven J 2006;2:e43.
- 58. Doughty JC, Anderson JH, Willmott N, McArdle CS. Intra-arterial administration of adriamycinloaded albumin microspheres for locally advanced breast cancer. Postgrad Med J 1995;71:47-9.
- 59. Hasemeier B, Christgen M, Kreipe H, Lehmann U. Reliable microRNA profiling in routinely processed formalin fixed paraffin-embedded breast cancer specimens using fluorescence labelled bead technology. BMC Biotechnology 2008;8:90.
- 60. Sabel MS, Skitzki J, Stoolman L, Egilmez NK, Mathiowitz E, Bailey N. Intratumoral IL-12 and TNF-

a–Loaded Microspheres Lead To Regression of Breast Cancer and Systemic Antitumor Immunity.

- 61. Chaurasia M, Chourasia MK, Jain NK, Jain A, Soni V, Gupta Y, et al. Cross-Linked Guar Gum Microspheres: A viable approach for improved delivery of anticancer drugs for the treatment of colorectal cancer. AAPS PharmSciTech 2006;7:74.
- 62. AlfL, HiromitsuY, HirofumiT, YoshiakiK. Microsphere design for the colonic delivery of 5fluorouracil. J Control Release 2003;90:313-22.
- 63. Blumen SR, Cheng K, Ramos-Nino ME, Taatjes DJ, Weiss DJ, Landry CC. Unique uptake of acidprepared mesoporous spheres by lung epithelial and mesothelioma cells. Am J Respir Cell Mol Biol 2007;36:333-42.
- 64. Chao P, Hu P, Stein S, Sinko P. Microsphere-based camptothecin passive lung targeting delivery system. Anti-cancer efficacy evaluation in an orthotopic lung cancer rat model. Pharmaceutical Science, Rutgers University Available from: http://www.aapsj. org/abstracts/AM_2005/AAPS2005-003046.pdf.
- Kumagai S, Sugiyama T, Nishida T, Ushijima K, Yakushiji M. Improvement of intraperitoneal chemotherapy for rat ovarian cancer using cisplatincontaining microspheres. Japanese J Cancer Res 1996;87:412-7.
- Jie Ma. Immunotherapy of Ovarian Cancer by Anti-CA125 Antibody. In: Bardos AP, Editor. Treatment of Ovarian Cancer. 1st ed. New York: Nova Science Publishers Inc; 2005. p. 79-86.
- 67. R T Liggins, T Cruz, W Min, L Liang, W L Hunter, H M Burt. Intra-articular treatment of arthritis with microsphere formulations of paclitaxel: biocompatibility and efficacy determinations in rabbits. Inflamm Res. 2004 Aug;53(8):363-72.
- Do, D., D'Souza, Enriquez, & Rizvi. Formulation and evaluation of drug-loaded targeted magnetic microspheres for cancer therapy. International Journal of Nanomedicine, 2013;1393
- 69. Francesco Blasi, Stefano Aliberti, Paolo Tarsia. Clinical applications of azithromycin microspheres in respiratory tract infections. Int J Nanomedicine. 2007;2;551-9.
- 70. Molday, R. S., Yen, S. P. S., & Rembaum, A. Application of magnetic microspheres in labelling and separation of cells. *Nature*, 1977;268, 437-438.
- Kang, M. L., Cho, C. S., & Yoo, H. S. Application of chitosan microspheres for nasal delivery of vaccines. *Biotechnology advances*, 2009;27;857-65.
- 72. Srivastava, P., & Visht, S. (2013). Application and advancement of microsphere as controlled delivery system: A review. *Int J Pharma Life Sci*, 203;4.