

# Design Development and Characterization of Clotrimazole Nano Sponge Gel

SRI CHARAN<sup>1</sup>, K.KISHORE<sup>2</sup>, G.VIJAY KUMAR<sup>3</sup>

Department Of Pharmacy, Kgr Institute Of Technology And Management Rampallyvill, Keesara Mdl, M.M Dist-5013, Ts

Received: 15.11.22, Revised: 19.12.22, Accepted: 07.01.23

## ABSTRACT

**Objectives:** Nanosponge is a new concept for the drug delivery system. In the present research work, an attempt was made to develop nanosponge based topical hydrogel containing Clotrimazole. Nanosponges are mainly used to provide the sustained release and thereby reduce the side effects caused by conventional dosage form.

**Methods:** Nanosponges were prepared using emulsion solvent diffusion method by using ethyl cellulose and eudragit S 100 in different concentrations. The prepared nanosponges were evaluated for preformulation parameters. The nanosponges formulation of all batches were evaluated for production yield, entrapment efficiency. Optimized formulation was evaluated for SEM analysis and incorporated into a gel base. The nanosponges containing gel was evaluated for pH determination, spreadability, swelling studies, viscosity determination, and in vitro diffusion study using franz diffusion cell for 10 hrs.

**Results and Discussion:** The results of FTIR analysis showed that there is no physical and chemical interaction between drug and other excipients. F2, F5 are considered to be the optimized nanosponge formulation. G1 was considered to be the best formulation. The data from the in vitro release study were fitted to various mathematical models. The results of mathematical model of fitting data obtained indicated the best fit in all cases. Stability study for 1 months at various condition shows G1 has good stability.

**Conclusion:** The study indicates that the rate of drug release can be improved by incorporating drug into the nanosponges and thus it can improve the targeting of the drug at the specific site and thereby reduce systemic toxicity.

**Keywords:** development, clotrimazole.

## INTRODUCTION

Nanosponges are tiny mesh – like nanoporous particular structure in which a large variety of substances can be encapsulated or suspended, and then be incorporated into a dosage form. They have a proven spherical colloidal nature, reported to have a very high solubilization capacity for poorly soluble drugs by their inclusion and non-inclusion behavior. Nanosponges have recently been developed and proposed for drug delivery. Nanosponges can solubilize poorly water soluble drug and provide prolonged release as well as improving drugs bioavailability. Nanosponges are able to load both hydrophilic and hydrophobic drug molecules because of their inner hydrophobic cavities and external hydrophilic branching, thereby offering unparalleled flexibility. Nanosponges are more like a three- dimensional network or scaffold. The backbone is a long length of polyester which is mixed in solution with small molecules called crosslinkers that act like tiny grappling hooks to fasten different parts of the polymer together. The nanosponges are encapsulating type of

nanoparticles which encapsulates the drug molecules within its core[4].

As the nanosponges have an open structure with pores on its surface ie; in the surrounding of nanosponges they do not have any uninterrupted membrane, the active substance is added to the vehicle in an encapsulated form. The encapsulated active substance is able to move freely from the particles into the vehicle until the vehicle gets saturated and the equilibrium is attained. When the product is applied on to the skin, the vehicle containing the active gets unsaturated causing a disturbance in the equilibrium. This will start a flow of the active from the sponge particle into the vehicle and from it to the skin until the vehicle is either dried or absorbed. Even after the withholding of the nanosponge particles on the surface of skin i.e. the stratum corneum, the release of active substance continues to skin for a long period of time.

For prolonged and controlled release of the drug products on the skin the nanosponges technology is the most efficient technology. Antifungal,

antibiotics, anti-inflammatory are the common type of drugs used in the topical application. Conventional products release the drug in a relatively high concentration this may lead to serious side effects but the nanosponge drug delivery system release the drug in a sustained and predictable manner. The nanosponges can be formulated into ointments, gels, creams, lotions[8]

### MATERIALS AND METHODS

Clotrimazole, Ethyl Cellulose, Eudragit S 100, Dichloromethane, Polyvinyl Alcohol, Carbopol 934, propylene glycol, Triethanolamine were obtained from Yarrow Chem products, Mumbai.

#### Formulation Of Nanosponges

and polyvinyl alcohol are used to prepare nanosponges. Two phases are used in this method— dispersed and continuous. The dispersed phase consists of ethyl cellulose and the drug, which is then dissolved in 20 ml of dichloromethane and some amount of polyvinyl alcohol (PVA) is added to 150 ml of the continuous phase (aqueous). Then, the mixture is stirred at the speed of 1000 rpm for about 2 h. The product i.e. the nanosponges are collected by filtration. Finally, the product is dried in an oven at a temperature of 40°C [Venkateshet al. Int J App Pharm, Vol 10, Issue 4, 2018, 1-5]

#### Materials used in the preparation of nanosponges

Polymer, Copolymer, - Hyper cross-linked polystyrenes, cyclodextrins and its derivatives like methyl  $\beta$ - cyclodextrine, 2-hydropropyl  $\beta$ - cyclodextrine. Ethyl cellulose (EC), polyvinyl alcohol (PVA), Crosslinkers like Di-phenyl Carbonate (DPC), diarylcarbonate, diisocyanates, pyromellitic anhydride, carbonyl diimidazole, 2,2-bis (acrylamide) acidic acid and dichloromethane. [8, 9]

#### Ultra-sound assisted synthesis

Polymers are made to react with crosslinkers in a flask without the solvent. The flask is placed in an ultrasound bath which is filled with water and heated up to 90°C and the mixture is sonicated for 5 h.

Then the mixture is cooled down to room temperature and then the product is broken into rough pieces. At last, the non-reacting polymer is removed by washing the product with water and refining is done using Soxhlet apparatus (ethanol) to obtain nanosponges[10].

#### Emulsion solvent diffusion method

In this method, different proportion or amount of ethyl cellulose and polyvinyl alcohol are used to prepare nanosponges. Two phases are used in this method— dispersed and continuous. The dispersed phase consists of ethyl cellulose and the drug, which is then dissolved in 20 ml of dichloromethane and some amount of polyvinyl alcohol (PVA) is added to 150 ml of the continuous phase (aqueous). Then, the mixture is stirred at the speed of 1000 rpm for about 2 h. The product i.e. the nanosponges are collected by filtration. Finally, the product is dried in an oven at a temperature of 40°C [11 Venkateshet al. Int J App Pharm, Vol 10, Issue 4, 2018, 1-5]

The nanosponges are prepared by emulsion solvent diffusion method. In this method two phases are used in different proportions. The dispersed phase having ethyl cellulose or eudragit S 100 and drug (Clotrimazole) get dissolved in dichloromethane (20 ml) and a definite amount of polyvinyl alcohol added to 100 ml of aqueous continuous phase. Then, the mixture was stirred properly at 1000 rpm for 2 hr. The required nanosponges were collected by the process of filtration by using membrane filter (pore size 0.45  $\mu$ m) and kept for drying in oven at 40°C for 24 hr. Nanosponge which are dried and stored in a desiccator are ensured of removal of residual solvents[16].

#### Composition Of Clotrimazole Nanosponges

**Table 1: Composition of Clotrimazole Nanosponges**

Ingredients	Formulations					
	F1	F2	F3	F4	F5	F6
Clotrimazole(mg)	100	100	100	100	100	100
Ethyl Cellulose(mg)	200	400	600			
Eudragit S 100(mg)				200	400	600
Dichloromethane(ml)	20	20	20	20	20	20
Polyvinyl alcohol(mg)	500	500	500	500	500	500
Distilled water(ml)	100	100	100	100	100	100

### Formulation Of Nanosponge Loaded Gel

The polymer Carbopol 934 was initially soaked in water for the gel for 2 hrs and dispersed by agitation at 600rpm by using magnetic stirrer to get smooth dispersion. Triethanolamine was

added to neutralise the pH. The previously prepared optimized nanosponge suspension was thereby added and permeation enhancers Propylene glycol was added as ethanolic solution to the aqueous dispersion[16]

**Table 2: Composition of nanosponges loaded gel**

Ingredients	Quantity
Clotrimazole loaded nanosponges(g)	1
Carbopol(g)	1
Propylene Glycol(ml)	5
Triethanolamine(ml)	q.s
Distilled water(ml)	100

### Evaluation Of Clotrimazole Loaded Nanosponges

#### Physical Examination

The prepared Clotrimazole loaded nanosponges were inspected visually for their colour and appearance

#### Production yield

The prepared nanosponges were collected and weighed Production yield of nanosponges was determined by formula mentioned below

$$\text{Production yield} = \frac{\text{Practical mass}}{\text{Theoretical mass}} \times 100$$

#### Surface Morphology

Scanning electron microscopy was used to analyze particle size and surface topography was operated at 15kV acceleration voltage. A concentrated aqueous suspension was spread over a slab and dried under vacuum. The sample was shadowed in a cathodic evaporator with a gold layer 20nm thick. Photographs were elaborated by an image processing program and individual diameters were measured to obtain mean particle size[16].

#### Entrapment Efficiency

To calculate the entrapment efficiency, accurately weighed quantity of nanosponges (10mg) with 5ml of methanolic HCl (HCl: Methanol-10:1) in a volumetric flask was shaken for 1min using vortex mixer. The volume was made upto 10ml with Methanolic HCl. Then the solution was filtered and diluted and the concentration of drug was determined spectrometrically at 295nm[16].

$$\text{Entrapment Efficiency} = \frac{\text{Actual drug content in nanosponges}}{\text{Theoretical drug content}} \times 100$$

### Evaluation Of Prepared Nanosponge Loaded Gel

#### Visual Inspection

The organoleptic properties, such as colour,

odour, homogeneity, and physical appearance of gel containing nanosponges were checked by visual inspection.

#### pH determination

The pH of the prepared nanosponge loaded hydrogel formulations were determined by using a digital pH meter. One gram of gel was dissolved in 100 ml of distilled water and stored for two hours. Then, pH measurement was performed. The measurement of pH of each formulation was done in triplicate and average values were calculated[16].

#### Viscosity Measurement

The viscosity of prepared hydrogels was measured using Brookfield viscometer. Viscosity was measured at 25°C at 100 rpm using spindle no.LV- 61[38].

#### Spreadability Studies

Spreadability is a term expressed to denote the extent of the area to which the gel readily spreads on application to the skin. The therapeutic efficacy of a semisolid formulation also depends on its spreading value. 1 g of the formulation was placed within a circle of 1cm diameter pre-marked on a ground glass slide. The gel formulation was sandwiched between this slide and the second slide having the same dimension. A weight of 500 g was allowed to rest on the upper glass slide for 5 min. The increase in the diameter due to gel spreading was noted. The spreadability was then calculated from the following formula[21].

$$S = M \times L/T$$

S= Spreadability M = Mass in grams

L=Length of the slide T =Time

#### Drug Content Estimation

1 g of prepared Clotrimazole nanosponge loaded hydrogel formulation containing drug equivalent to 100 mg was extracted with 30 ml of ethanol.

The volume was made up to 100 ml with phosphate buffer 7.4. The solution was filtered. The absorbance of the resulting solution was measured at 295 nm using a UV spectrophotometer after suitable dilutions. The drug content of the formulation was determined using the following equation.

$$\% \text{ Drug content} = \frac{\text{Actual concentration of drug in the formulation}}{\text{Theoretical concentration of drug}} \times 100$$

### In Vitro Drug Release Studies

*In vitro* release study of Clotrimazole nanosponges loaded hydrogel was carried out by using Franz diffusion cell. The formulation was taken in the donor compartment and phosphate buffer saline was taken in the receptor compartment. The cellophane membrane previously soaked overnight in the diffusion medium (phosphate buffer 7.4) was placed between the donor and the receptor compartment. 1 g of the formulation was spread uniformly on the cellophane membrane, which is in contact with the receptor medium. The whole assembly was placed on the thermostatically controlled magnetic stirrer with continuous stirring and the temperature of the medium was maintained at  $37 \pm 0.5^\circ\text{C}$ . At specific intervals, 1 ml of sample was withdrawn from the receptor compartment and replaced with an equal

volume of Phosphate buffer 7.4. After suitable dilutions, the absorbance of the sample was determined at 295 nm by UV-visible spectrophotometer[21].

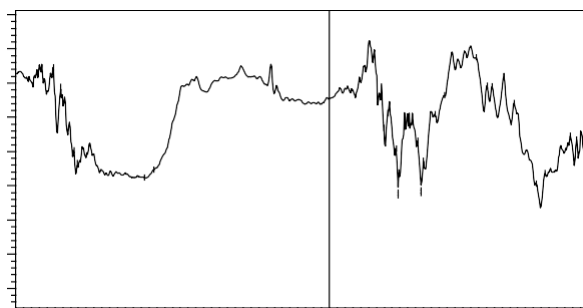
### Drug Release Kinetics

Release kinetics of drug from the dosage form was determined by various mathematical models such as zero order, first order, korsmeyer-peppas and higuchi model.

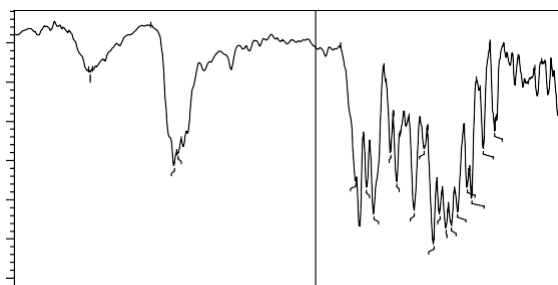
- Zero Order Plots(Cumulative percentage drug released) v/s time
- First Order Plots(Log cumulative percent drug remaining) v/s time
- Higuchi Plots(Cumulative percentage drug release) v/s square root of time
- Korsmeyer-Peppas Plots(Log cumulative percentage drug release) v/s log time

### Results And Discussions

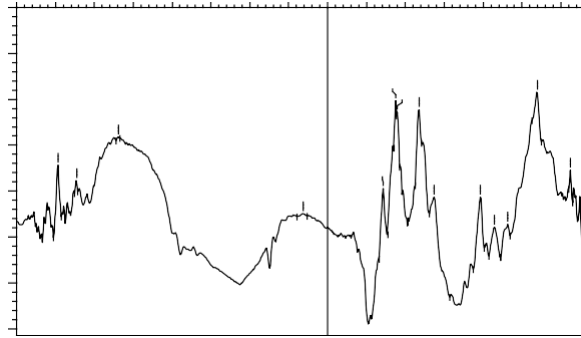
FTIR analysis was carried out for pure drug and drug excipient mixtures. Spectrum of drug showed the prominent peaks with respect to functional groups. The spectrum of physical mixture of drug with excipients showed that there is no significant interaction between the drug, polymer and excipients. In the spectrum of drug polymer mixtures the characteristic peak of drug was not altered.



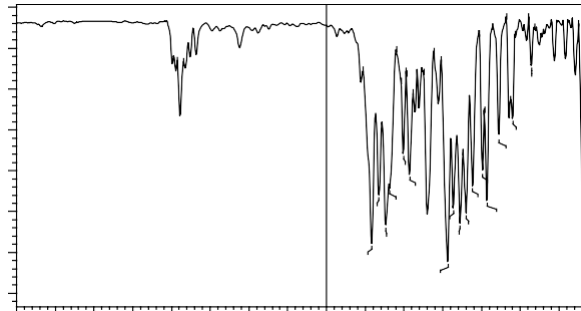
**Fig 1: FTIR Spectrum of the Clotrimazole**



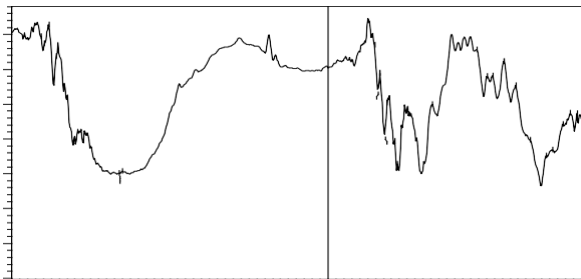
**Fig 2: FTIR Spectrum of Clotrimazole and Ethyl Cellulose**



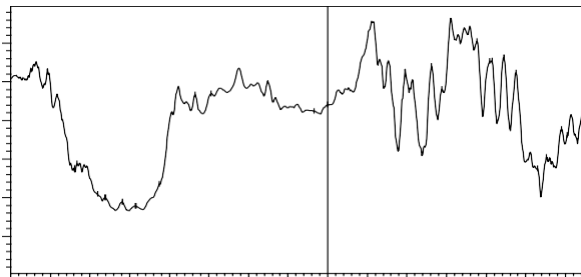
**Fig 3: FTIR Spectrum of Clotrimazole and Eudragit S 100**



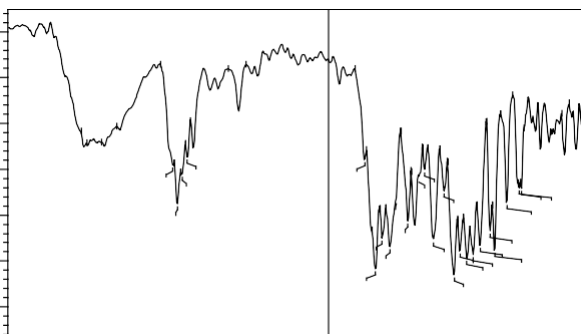
**Fig 4: FTIR Spectrum of Clotrimazole and Dichloromethane**



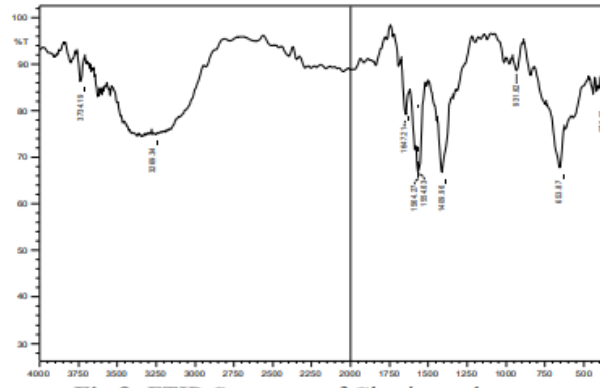
**Fig 5: FTIR Spectrum of Clotrimazole and Polyvinyl Alcohol**



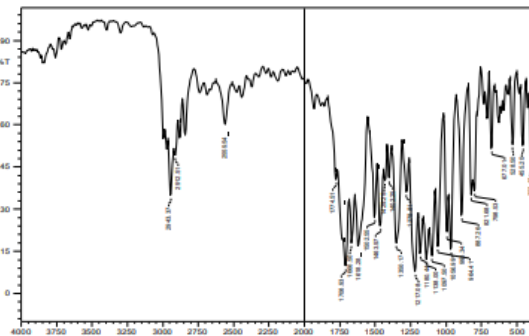
**Fig 6: FTIR Spectrum of Clotrimazole and Carbopol 934**



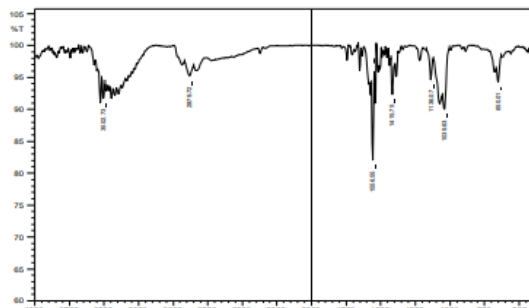
**Fig 7: FTIR Spectrum of Clotrimazole and Ethanol**



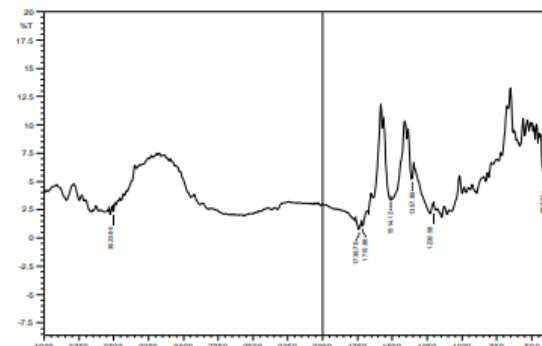
**Fig 8:** FTIR Spectrum of Clotrimazole and Triethanolamine



**Fig 9:** FTIR Spectrum of Clotrimazole and Propylene Glycol



**Fig 10:** FTIR Spectrum of Clotrimazole and Mixture 1



**Fig 11:** FTIR Spectrum of Clotrimazole and Mixture 2

## Formulation Development



**Fig 12: F2 Formulation**



**F13: G1 Formulation**



**Fig 14: F5 formulation**



**Fig 15: G2 formulation**

### Evaluation Of Clotrimazole Loaded Nanosponge Physical Examination

Physical evaluation of nanosponges were shown in the table c. From the physical evaluation of all the

batches formulated. It was concluded that the nanosponges of all batches had desirable physical properties.

**Table 3: Physical Examination**

Sl. No	Formulation Code	Colour	Appearance
1	F1	White	Powder
2	F2	White	Powder
3	F3	White	Powder
4	F4	White	Powder
5	F5	White	Powder
6	F6	White	Powder

**Production Yield**

The production yield was calculated. Production yield of nanosponges are shown in the table d. The production yield of the prepared nanosponges of

Clotrimazole ranges from of 74% to 89%. It revealed that all formulation have good production yield.

**Table 4: Production Yield**

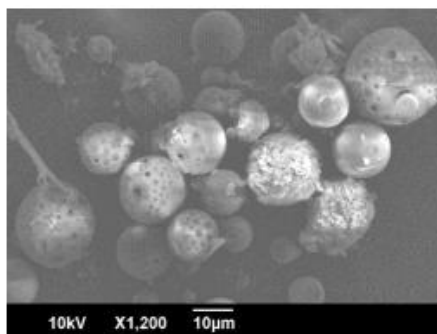
Sl No	Formulation code	Production yield (%)
1	F1	77
2	F2	85
3	F3	87
4	F4	74
5	F5	82
6	F6	89

**Entrapment Efficiency**

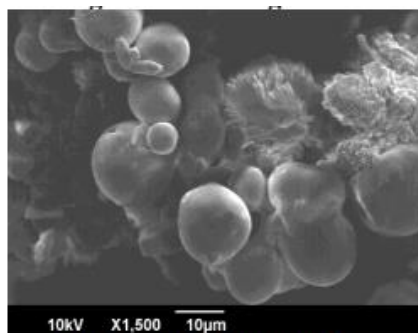
The entrapment efficiency of all batches were tested. The results were shown in the table e. The

results show that the entrapment efficiency were in the range of  $62.9 \pm 1.55\%$  to  $87.7 \pm 1.16\%$ . The entrapment efficiency was higher in F2 and F5.

**Surface Morphology**



**Fig 16: SEM image of F2**



**Fig 17: SEM image of F5**



**Entrapment Efficiency**

The entrapment efficiency of all batches were tested. The results were shown in the table e. The

results show that the entrapment efficiency were in the range of  $62.9 \pm 1.55\%$  to  $87.7 \pm 1.16\%$ . The entrapment efficiency was higher in F2 and F5

**Table 5: Entrapment Efficiency**

SI No	mulation Code	Entrapment Efficiency(%)(*±SD)
1	F1	$81.8 \pm 1.77$
2	F2	$87.7 \pm 1.16$
3	F3	$68.66 \pm 0.84$
4	F4	$62.9 \pm 1.55$
5	F5	$78.13 \pm 0.65$
6	F6	$66.3 \pm 0.57$

\*Average of three determinants, SD=Standard deviation

**Evaluation Of Prepared Nanosponge Loaded Gel**

G1: Gel containing F2 formulation  
G2: Gel containing F5 formulation

**Visual Inspection**

**Table 6: Visual Inspection**

Formulation Code	G1	G2
Colour	White	White
Odour	Odourless	Odourless
Appearance	Transparent	Transparent
Homogeneity	Homogeneous	Homogeneous

**Ph Determination**

The pH determination was done and the results were shown in the table g. The pH of the

formulations were found to be satisfactory. The pH of G1 was found to be  $7.02 \pm 0.29$  and the pH of G2 was found to be  $6.76 \pm 0.16$ .

**Table 7: pH Determination**

SI No	Formulation code	pH (*±SD)
1	G1	$7.02 \pm 0.29$
2	G2	$6.76 \pm 0.16$

\*Average of three determinants, SD=Standard deviation

**Viscosity Measurement**

The viscosity of the gel was determined. The viscosity was measured by the Brookfield viscometer spindle no. 61 at 100rpm. The result

was shown in the table h. The viscosity of G1 and G2 was found to be 7285 centipoise and 8154 centipoise respectively.

**Table 8: Viscosity Measurement**

SI No	mulation code	Viscosity (cps) (*± SD)
1	F1	$7285 \pm 0.32$
2	F2	$8154 \pm 0.26$

\*Average of three determinants, SD=Standard deviation

**Spreadability**

The spreadability of the formulations were done and the result was shown in the table i.

Spreadability of G1 and G2 was found to be  $7.56 \text{ gm-cm/s}$  and  $6.24 \text{ gm-cm/s}$  respectively.

**Table 9: Spreadability**

Sl No	Formulation Code	readability (gm-cm/s) (*± SD)
1	G1	7.56±0.17
2	G2	6.24±0.21

\*Average of three determinants, SD=Standard deviation

**Drug Content**

Drug content was calculated and the results were

shown in the table j. G1 shows high drug content 98.09%.

**Table 10: Drug Content Fig 18: In vitro drug release of Gel 1 and Gel**

Sl. No	Formulation code	rug content (*± SD)
1	G1	98.09±0.13
2	G2	95.27±0.53

\*Average of three determinants, SD=Standard deviation

**In Vitro Drug Release of G1 And G2**

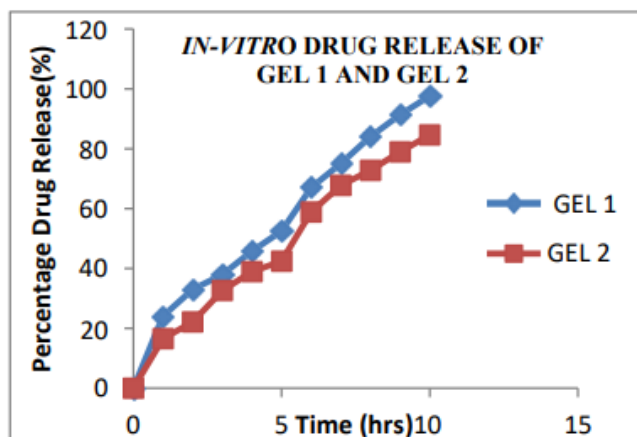
The in vitro drug release of the nanosponges loaded gel was carried franz diffusion cell apparatus with phosphate buffer 7.4 for 10 hrs the

results were shown in the table k. The plot of percentage drug release v/s time (hrs) was shown in figure 18. Gel 1 shows high percentage drug release.

**Table 11: Percentage Drug Release**

Sl.No	TIME (h)	CENTAGE OF DRUG RELEASE (*± SD)	
		Gel 1	Gel 2
0	0	0	0
1	1	24.08 ± 0.50	16.54±0.74
2	2	32.96±1.00	22.13±1.69
3	3	38 ±0.36	32.65±0.96
4	4	45.73± 0.65	38.9±0.31
5	5	52.46 ±1.60	42.43±1.51
6	6	67.06± 1.11	58.87±1.01
7	7	75 ± 1.11	67.76±1.62
8	8	84 ± 1.22	72.76±1.63
9	9	91± 1.24	78.9±1.69
10	10	97.6± 0.46	84.6±1.24

\*Average of three determinants, SD=Standard deviation



**Fig 18: In vitro drug release of Gel 1 and Gel**

Swelling Studies

**Table 12: Swelling Studies**

Sl. No	Formulation code	Percentage Swelling (%)
1	G1	74%
2	G2	68%

Swelling study was performed and the results were shown table I. From analyzing the percentage swelling, we can conclude that G1 shows high swelling percentage 74%.

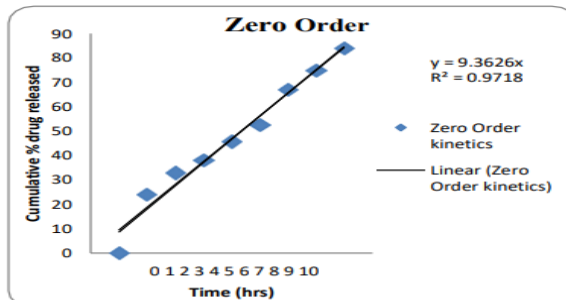
first order, Higuchi model and Korsmeyer peppas model.

**Kinetic Model Gel 1**

The diffusion profile of optimized formulation G1 was fitted to various kinetic models like zero order,

**Zero Order Plot**

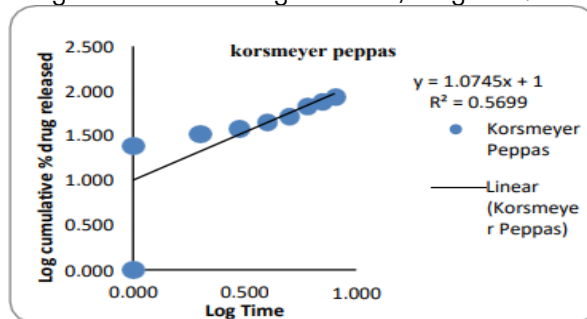
Graph was plotted between cumulative % drug released v/s time.



**Fig 19:** Zero order plot for drug release kinetics of G1 formulation

**Korsmeyer peppas plot**

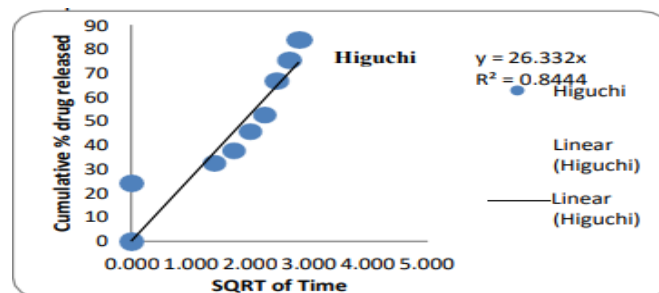
Graph was plotted between log cumulative % drug release v/s log time.



**Fig 20:** Korsmeyer peppas plot for drug kinetics of G1 Formulation

**Higuchi plot**

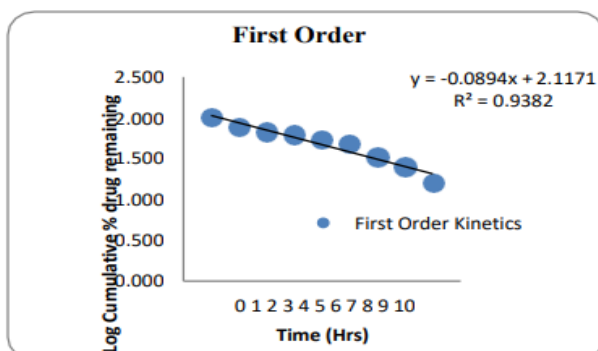
Graph was plotted between % cumulative drug released v/s square root of time



**Fig 21:** Higuchi plot for drug release kinetics of G1 Formulation

**First order plot**

Graph was plotted between log cumulative % drug remaining v/s time



**Fig 22:** First order plot for drug release kinetics of G1 formulation

**Table 13:** R2 Values of various kinetics release data of optimized nanosponge gel

G1	Zero order	First order	Korsmeyer-peppas model	Higuchi model
R <sup>2</sup>	0.975	0.938	0.532	0.844

The diffusion profile of optimized formulation G1 was fitted to zero order, first order, Higuchi model and Korsmeyer Peppas model to ascertain the kinetic modelling of the drug releasing mechanism showed in figure 19-22. The correlation coefficient (R<sup>2</sup>) for all the formulations using different kinetic equation is listed in table m. It was found that the in vitro drug release of optimized formulation G1 was best explained by zero order as the plot show highest linearity (R<sup>2</sup>= 0.975) followed by first order. (R<sup>2</sup> =0.938) The R<sup>2</sup> was used to accuracy

of fit. The formulation G1 provide best fit to the zero order model.

**Stability Studies**

Stability studies were performed as per ICH guidelines. The formulation G1 was selected for the stability studies. After 1 month storage the nanosponge loaded gel were evaluated for various parameters like physical appearance, pH, drug content, percentage drug release.

**Table 14:** Stability study data at 5 0C±20C

Days	pH	Physical stability	Drug content	Percentage of drug release
0	7.02±0.29	No Change in appearance	98.09±0.13	97.6± 0.46
15	7.11±0.14	No change in appearance	97.45±0.23	96.51±1.13
30	6.93±0.27	No change in appearance	96.14±0.34	96.34±1.21

**Table 15:** Stability study data at 25<sup>0</sup>C±2<sup>0</sup>C

Days	pH	Physical stability	Drug content	Percentage of drug release
0	7.02±0.29	No Change in appearance	98.09±0.13	97.6± 0.46
15	7.22±1.12	No change in appearance	97.25±0.18	96.45±1.18
30	6.95±0.29	No change in appearance	96.13±0.19	96.30±1.10

Formulation G1 after 30 days stability study at different conditions shows that there is no major change in the formulation after the storage as initial. The study shows no major difference was found before and after the storage and all are in satisfactory range. Therefore formulation remains stable for sufficient range. Therefore formulation remains stable for sufficient time after the storage of 30 days.

**CONCLUSION**

In the present study, nanosponge formulation were presented as a new attempt to enhance the bioavailability of the drug Clotrimazole there by provide a sustained delivery to the targeted site for the treatment of several fungal diseases. Nanosponge was prepared by emulsion solvent diffusion method using ethyl cellulose and eudragit S 100 at different concentration. FTIR studies showed that absence of incompatibility

between drug and excipients. Formulation F2 and F5 was found to be the best formulation based on the entrapment efficiency. F2 and F5 were selected to formulate as gel, G1 and G2 respectively. G1 was found to be the best formulation based on swelling study(74%), viscosity ( $7285 \pm 0.32$  cps), spreadability (7.56 gm-cm/s), drug content( $98.09 \pm 0.13\%$ ) and in vitro study shows 97.6% of drug release at 10th hour of study. In vitro follows zero order kinetics in drug release kinetic analysis. The optimized formulation G1 was found to be stable during stability study.

## REFERENCES

- Shivani S, Poladi KK. Nanosponges-novel emerging drug delivery system- a review. *International journal of pharmaceutical sciences and research*. 2015;6(2):529-40.
- Swetha T, Chakraborty T. Nanosponges- new colloidal drug delivery system for topical drug delivery. *Indo American journal of pharmaceutical sciences*. 2019; 6 (2): 4263-76.
- Bowmik H, Venkatesh ND, Kuila A, Kumar KH. Nanosponges: a review. *International journal of applied pharmaceuticals*. 2018;10(4) :1-5.
- Shringirishi M, Prajapati SK, Mahor A, Alok S, Yadav P, Verma A. Nanosponges: a potential nanocarrier for novel drug delivery- a review. *Asian Pacific Journal of Tropical Disease*. 2014;4(2):519-26.
- Salunkhe A, Kadam S, Magar S, Dangare K. Nanosponges: a modern formulation approach in drug delivery system. *World Journal of Pharmacy and Pharmaceutical Sciences*. 2018;7(2):575-92.
- Pawar YA, Naik AK, Jadhav KR. Nanosponges: a novel drug delivery system. *Asian journal of pharmaceuticals*. 2016;10(4):456-61.
- Tiwari H, Mahor A, Dixit ND, Kushwaha M. A review on nanosponges. *World Journal of Pharmacy and Pharmaceutical Sciences*. 2014;3(11):219-33.
- Subramanian S, Singireddy A, Krishnamoorthy K, Rajappan M. Nanosponges: a novel class of drug delivery system – review. *J Pharm Pharmaceut Sc*. 2012;15(1):103 – 11.
- Nasir A, Kousar P, Amjad H, Sumera L, Shaiq U, Pervaiz A, Shah M A. Nanosponge-based hydrogel preparation of fluconazole for improved topical delivery. *Tropical Journal of Pharmaceutical Research*. 2019;18(2):215-22.
- Osmani RA, Aloorkar NH, Kulkarni AS, Kulkarni PK, Hani U, Thirumaleshwar S, et al. Novel cream containing microsponges of anti- acne agent: formulation development and evaluation. *Curr Drug Deliv*. 2015;12:504-16.
- Swaminathan S, Cavalli R, Trotta F. Cyclodextrin-based nanosponges: a versatile platform for cancer nanotherapeutics development. *Wiley Interdiscip Rev Nanomed Nanobiotechnol*. 2016;8:579-601.
- Pushpalatha R, Selvamuthukumar S, Kilimozhi D. Nanocarrier mediated combination drug delivery for chemotherapy – A review. *J Drug Deliv Sci Technol*. 2017;39:362-71.
- Momin MM, Zaheer Z, Zainuddin R, Sangshetti JN. Extended release delivery of erlotinib glutathione nanosponge for targeting lung cancer. *Artif Cells Nanomed Biotechnol*. 2018;46:1064-75.
- Muralidharan S, Venugopal V, Kumar J, Joshua H O. Bioanalytical method and validation of Clotrimazole nanoparticles using HPLC method. 2015;4(7):384-8.
- Bioavailability of micronized Clotrimazole from corn oil in water, emulsion, aqueous suspension and commercial tablet dosage formulation. *Journal of pharmaceutical sciences*. 2018;64(5): 234-45.
- Prathima S, Sreeja k. Formulation and evaluation of voriconazole loaded nanosponges for oral and topical delivery. *International journal of drug development and research*. 2012;5(1):55-69.
- Abbas N, Hussain A, Hatizma, Perveen, K. Formulation and evaluation of fluconazole loaded nanosponges for improved topical drug delivery. *British Journal of Pharmacy*. 2017;5(8):41-2.
- Kumar SK, Hematheerthani N, Ratna VJ, Saikishore V. Formulation and evaluation of miconazole nitrate loaded nanosponges for vaginal drug delivery. *Indo American Journal of American Journal*. 2015; 2(6):1028-37.
- Reddy N, Stella P, Lavanya A, Kumar U, Priyanka. Fabrication and characterization of itraconazole loaded nanosponge gel. *World Journal of Pharmaceutical Research*. 2019;8(5):1184-204.
- Anjali SK, Sheri PS, Kuriachan MA. Formulation and evaluation of antifungal nanosponge loaded hydrogel for topical delivery. *International journal of pharmacy and pharmaceutical research*. 2018;13(1):363-79.
- Raja NV, Kumar G, Anusha K. Fabrication and evaluation of ciprofloxacin loaded nanosponges for sustained release. *International Journal of Research In Pharmaceutical and Nano Sciences*. 2013;2(1):1-9.
- Khalid A, Ansari, Torne JS, Pradeep R, Valvia, Tortia F. Paclitaxel loaded nanosponges; in vitro characterization and cytotoxicity study on MCF - 7 cell line culture. *Current drug delivery*. 2011;8(2):194-202.
- Seema G, Kumar SA, Manoj B. Development and evaluation of Curcumin loaded nanosponges for colon drug delivery. *World Journal of Pharmaceutical Research*. 2015;4(5):1650-66.
- Monica RP, Rohini C, Bhingole. Developed and evaluated the nanosponge- based pediatric-controlled release dry suspension of gabapentin for reconstitution. *Drug development and*

- Industrial Pharmacy. 2015;6(8):356-69.
25. Aldaswari H, Badr-Eldin, Labib GS, Kamel A. Design and formulation of a topical hydrogel integrating lemongrass loaded nanosponges with an enhanced antifungal effect: in vitro: invivo evaluation. *International journal of nanomedicine*. 2015;10(5): 893-902.
  26. Swaminathan S, Vavia PR, Trotta F, Torne S. Formulation of beta cyclodextrin based nanosponges of itraconazole. *J Incl Phenom Macrocycl Chem*. 2007;5(7):89-94.
  27. Darshini M, Katamreddy JD, Shilpaja, Umasakar K. Atorvastatin loaded nanosponges- a novel strategic approach for enhanced bioavailability. *World Journal of pharmacy and pharmaceutical Sciences*. 2013;6647(8): 1223-36.
  28. Pavani A, Rama B. Formulation and invitro characterization of flurbiprofen nanosponges. *International Journal of Research in pharmacy and chemistry*. 2018;8(4):577-82.
  29. Padya K, Shah.N, Gohil.DX, Seth AK. Development of resedronate sodium loaded nanosponges by experimental design: optimization and in vitro characterization. 2019;8(12):523-43
  30. Panjuri SC, Ravouru N, Damineni S, Sailakshmi BN, Poreddy SR. Formulation and Evaluation of Lanzoprazole loaded nanosponges. *Turk. J .Pharmsci*. 2016;13(3):304-10.
  31. *Indian Pharmacopoeia*. 2018:2195-97.
  32. [www.health.com](http://www.health.com)
  33. [www.medicines.org.uk](http://www.medicines.org.uk)
  34. Dash AK, Mishra D. Development ,validation and stability study of Clotrimazole in bulk and pharmaceutical dosage form by UV spectroscopic method. *Asian Journal of Pharmaceutical research*. 2012;2(1)66-69
  35. Cavalli R, Trotta F, Tumiatti W. Cyclodextrin-based nanosponges for drug delivery. *Journal of Inclusion Phenomena and Macro Chemistry*. 2012;56(1-2): 209-13.
  36. Vyas SP, Khar RK. Targeted and controlled drug delivery- novel carrier systems. *Molecular basis of targeted drug delivery*. CBS Publishers and Distributors; New Delhi. 2008;38- 40.
  37. Aldawsari H M, Badar-Eldin S M, Labib G S and El-Kamel A H. Design and formulation of topical hydrogel integrating lemongrass loaded nanosponges with an enhanced antifungal effect: in vitro/in vivo evaluation. *International journal of nanomedicine*. 2015;10:893-902.
  38. Sharma R, Roderick B. Walker, Kamla P. Evaluation of kinetics and mechanism of drug release from econazole nitrate nanosponge loaded carbapol hydrogel. *Ind J Pham Edu Res*. 2011;45(1):25-31.
  39. Jilsha G, Vidya V. Nanosponge loaded hydrogel of cephalixin for topical delivery. *International Journal of Pharmaceutical Sciences and Research*. 2017;6(4):1200-34.
  40. Prathima S, Jahnavi AR. Formulation and evaluation of isoniazid loaded nanosponges for topical delivery. *Pharmaceutical Technology*. 2015;3(1):68-76.
  41. Nacht S, Kantz M. The microsponge: a novel topical programmable delivery system, in: *Topical Drug Delivery Systems*. David WO, Anfon H A editors; New York: Marcel Dekker. 1992;42:299-325.
  42. Eki S, Lei T, Jingquan L, Zhongfan J, Cyrille B, Thomas PD. Biodegradable star polymers functionalized with  $\beta$ -cyclodextrin inclusion complexes. *Bio macromolecules*.2009;10(9):2699- 700.
  43. Jenny A, Merima P, Alberto F, Francesco T. Role of  $\beta$ - cyclodextrin nanosponges in propylene photooxidation. *Carbohydrate Polymers*. 2011; 8(6):127-35.
  44. Aritomi H, Yamasaki Y, Yamada K, Honda H, Khoshi M. Development of sustained release formulation of chlorpheniramine maleate using powder coated microsponges prepared by dry impact blending method. *Journal of Pharma Sci and Tech*. 1996;56(1):49-56.
  45. Selvamuthukumar, Subramanian. Nanosponges: a novel class of drug delivery system – review. *J Pharm Pharma Sci*. 2012;15(1):103-111. 46. Kerali SV, Machevas P. Van P A, Shah VP. Bioavailability and bioequivalence: focus on physiological factors and variability. *Pharmaceutical research*. 2008; 1956-62.
  47. Yurtdas G, Demirel M, Genc L. Inclusion complexes of fluconazole with bcyclodextrin: physicochemical characterization and in vitro evaluation of its formulation. *J. Incl. Phenom. Macrocycl. Chem*. 2011;70:429-35.
  48. Mishra MK, Shikhri M, Sharma R, Goojar MP. Optimization, formulation, development and characterization of eudragit rs 100 loaded microsponges and subsequent colonic delivery. *International Journal of Drug Discovery and herbal Research*. 2011;1(1):8-13.
  49. Lala R, Thorat A, Gargote C. Current trends in  $\beta$ - cyclodextrin based drug delivery systems. *Int J Res Ayur Pharm*. 2011;2(5):1520-6.
  50. Shankar S, Linda P, Loredana S, Francesco T, Pradeep V, Dino A, Michele T, Gianpaolo Z, Roberta C. Cyclodextrin-based nanosponges encapsulating camptothecin: Physicochemical characterization, stability and cytotoxicity. *Eur J Pharm Biopharm*. 2010;74:193-201.
  51. Amber V, Shailendra S, Swarnalatha S: Cyclodextrin based novel drug delivery systems. *J pharmaceut sci*. 2012; 62:23-42.
  52. Rajeswari C, Alka A, Javed A, Khar RK: Cyclodextrins in drug delivery: an update review. *AAPS Pharm Sci Tech*. 2013; 6(2):329-57.
  53. Ramnik S, Nitin B, Jyotsana M, Horemam SN. Characterization of cyclodextrin inclusion

- complexes –a review. *J Pharm Sci Tech.* 2010;2(3):171-83.
54. Aithal KS, Udupa N, Srinivasan KK. Physicochemical properties of drug cyclodextrin complexes. *Indian drugs.* 2008; 32:293-305. 55. pubchem.ncbi.nlm.nih.gov
  56. www.sigmaldrich.com
  57. Duchene D, Vaution C, Glomot F. Cyclodextrin, their value in pharmaceutical technology. *Drug Dev Ind Pharm.* 2012;12(1113):2193-215.
  58. Torkaman M, Zare F. The use of ethyl cellulose polymer to control drug release of hydrocortisone acetate. *Oriental journal of chemistry.* 2017;33(4):1-10.
  59. Tayade PT, Vavia PP. Inclusion complexes of ketoprofen with  $\beta$ - cyclodextrins: Oral pharmacokinetics of ketoprofen in human. *Indian J Pharm Sci.* 2012; 68(2):164-70.
  60. Renuka S, Kamla P. Polymeric Nanosponges as an alternative carrier for improved retention of econazole nitrate onto the skin through topical hydrogel formulation. *Pharm Dev Technol.* 2011;16(4):367-76.
  61. Renuka S, Roderick BW, Kamla P. Evaluation of the kinetics and mechanism of drug release from econazole nitrate nanosponge loaded carbapol hydrogel. *Ind J Pharm Edu Res.* 2011;45(1):25-31.
  62. Isabelle A, Christine V, Helene C, Elias F, Patrick C. Sponge like Alginate nanoparticles as a new potential system for the delivery of antisense oligonucleotides. *antisense and nucleic acid drug development.* 2012;9:301-12.
  63. Rosalba M, Roberta C, Roberto F, Chiara D, Piergiorgio P, Leigh E, Li S, Roberto P. Antitumor activity of nanosponge- encapsulate camptothecin in human prostate tumors. *Cancer Res.* 2011;71:31-41.
  64. Torne SJ, Ansari KA, Vavia PR, Trotta F, Cavalli R. Enhanced oral paclitaxel bioavailability after administration of paclitaxel loaded nanosponges. *Drug Delivery.* 2010;17(6):419–25.
  65. Ansari KA, Torne SJ, Vavia PR, Trotta F, Cavalli R. Paclitaxel loaded nanosponges: in-vitro characterization and cytotoxicity study on MCF-7 cell line culture. *Curr Drug Deliv.* 2011;8(2):194-202.
  66. Shankar S, Vavia PR, Francesco T, Satyen T. Formulation of betacyclodextrin based nanosponges of itraconazole. *J Incl Phenom Macrocycl Chem.* 2007;57: 89–94.
  67. Swaminathan S, Cavalli R, Trotta F and Vavia PR. In vitro release modulation and conformational stabilization of a model protein using swellable polyamidoamine nanosponges of cyclodextrin. *J Incl Phenom Macrocycl Chem.* 2010;9765-9.
  68. Rao MR, Bajaj AN, Pardeshi AA, Aghav SS. Investigation of nanoporous colloidal carrier for solubility enhancement of